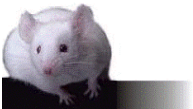


Superprimers

A New Approach for Genotyping Degraded DNA

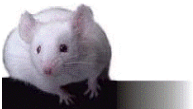


Superprimers

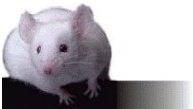
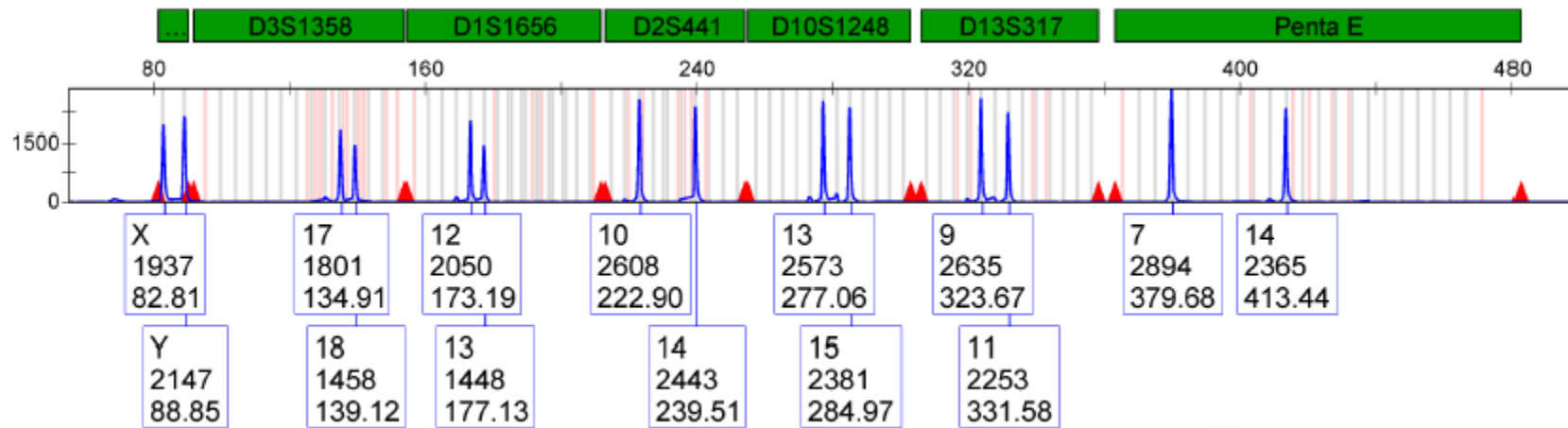
A New Approach for Genotyping Degraded DNA



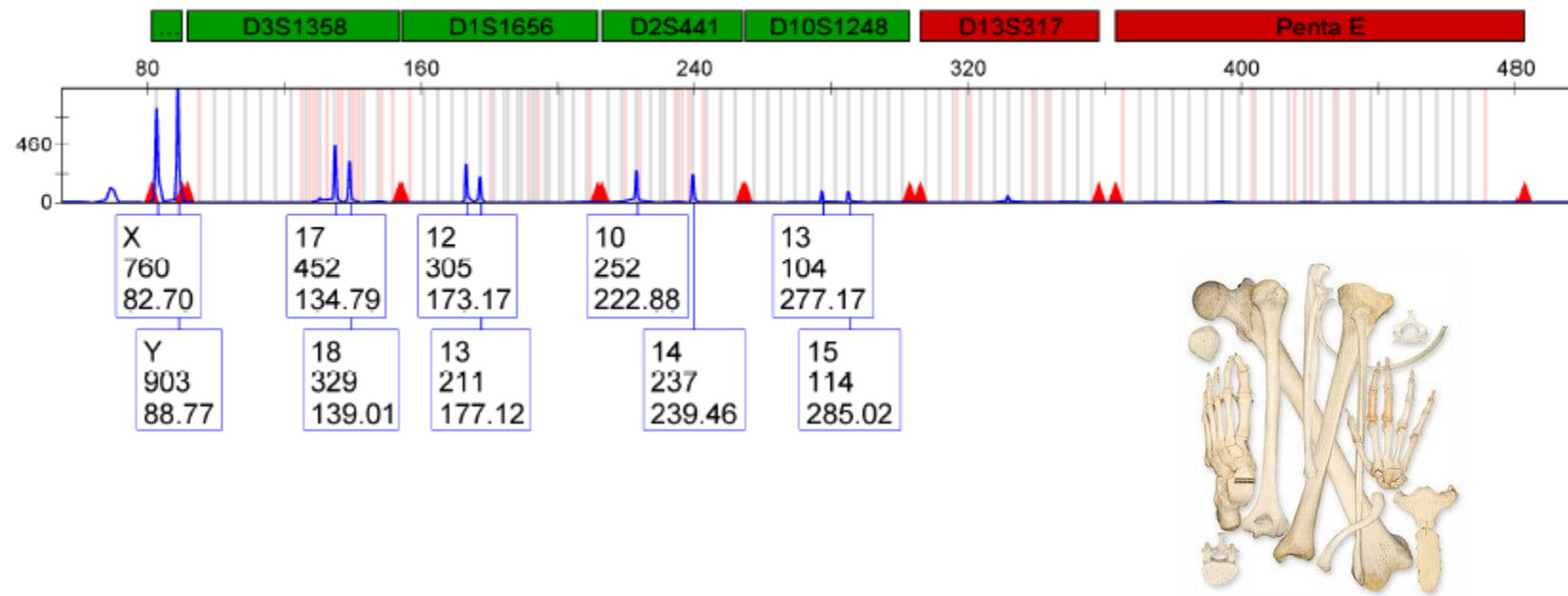
5'ccggaggtaaagggtgtcttaaagtgagaaagaataactgcatcttaacctattgggaggtcattgtaaagaggagagtgatggggtcagattgtacagagg
aggcacttcgtggtggtcaggagcacacactccagggcagtggtccaacctgagtctgccaggactagcaggttgctaaccacccctgtgtccagttt 3'



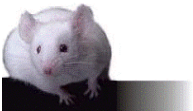
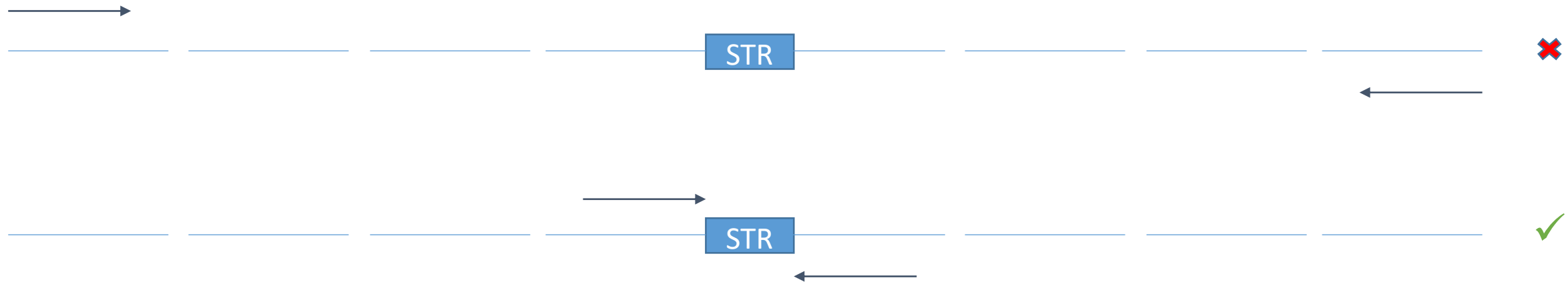
Current multiplex STR kit on intact DNA



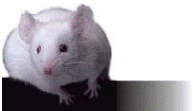
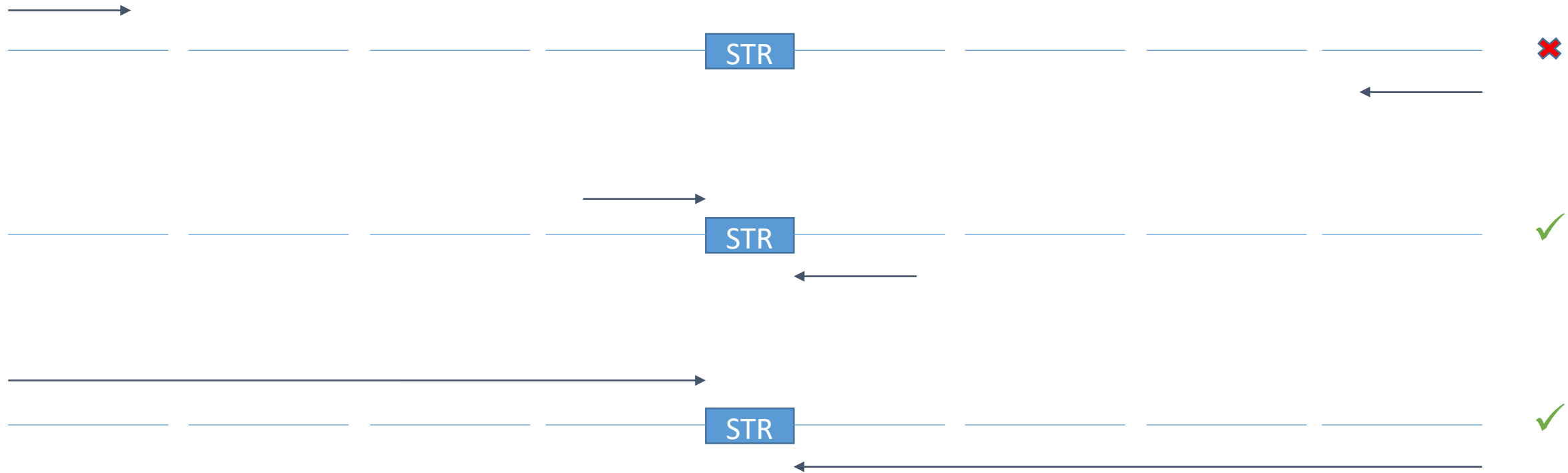
Current multiplex STR kit on degraded DNA



Superprimers on degraded DNA



Superprimers on degraded DNA



Amplifying with superprimers

120nt

ccggaggtaa aggtgtctta aagtgagaaa gaataactgc atcttaacct attgggaggt cattgtaaag aggagagtga tggggtcaga ttgtacagag gaggcacttc gtggtggtca

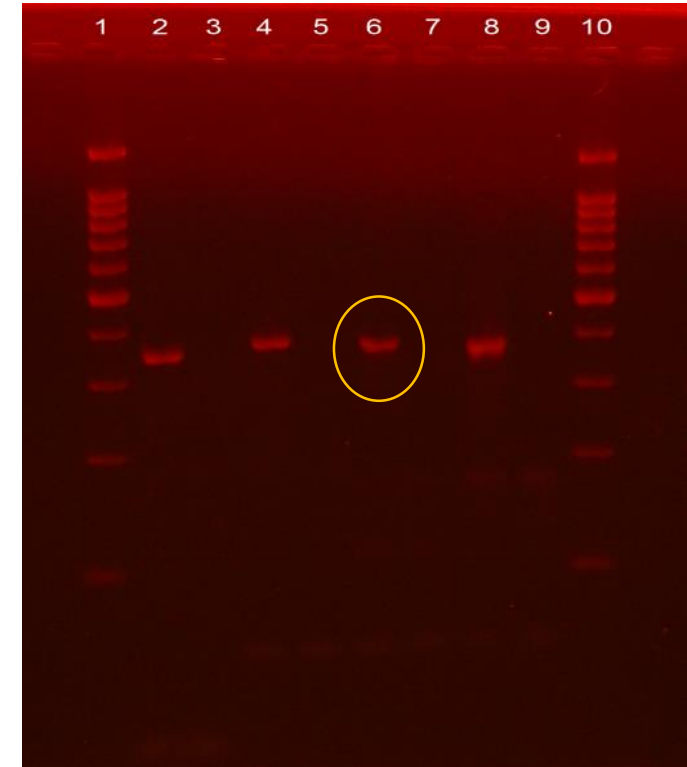
CSFFW2416HS (24nt) →

CSFFW120 (120nt) →

CSFFW200 (200nt) →

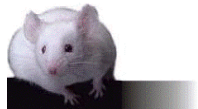
CSFFW260 (260nt) →

CSFFW300 (300nt) →



1: 100bp
4: CSFFW2416HS + CSFRV60J
8: CSFFW200 + CSFRV60J

2: CSFFW2416HSJ + CSFRV2216HS
6: CSFFW120 + CSFRV60J
10: 100bp



Biodynamics

AGAT

← CSFRV60J (60nt)

← CSFRV2216HS (22nt)

Amplifying with superprimers

200nt

ccggaggtaa aggtgtctta aagtgagaaa gaataactgc atcttaacct attgggaggt cattgtaaag aggagagtga tggggtcaga ttgtacagag gaggcacttc gtggtggtca
ggagcacaca ctccagggca gtgtccaac ctgagtctgc caaggactag caggttgcta accaccctgt gtctcagttt

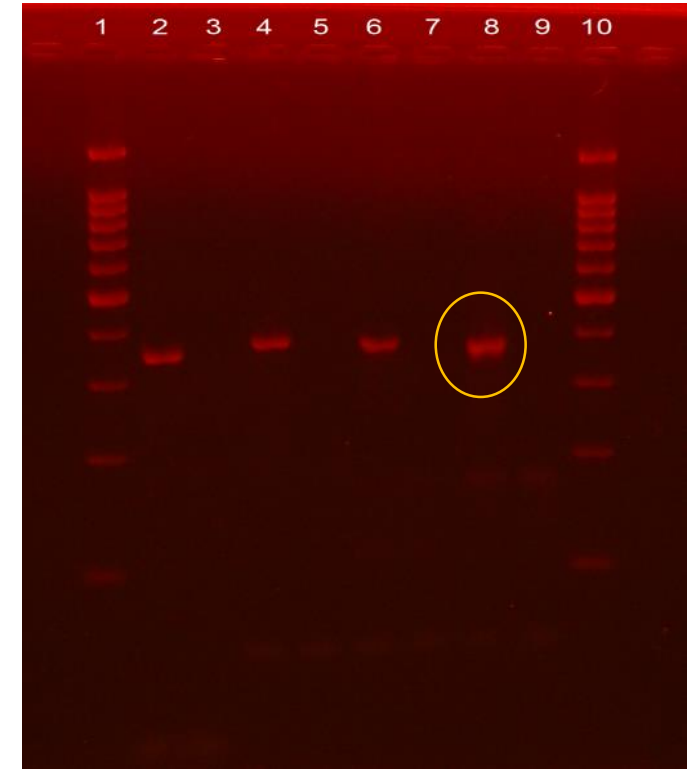
CSFFW2416HS (24nt) →

CSFFW120 (120nt) →

CSFFW200 (200nt) →

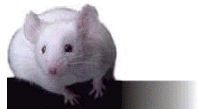
CSFFW260 (260nt) →

CSFFW300 (300nt) →



1: 100bp
4: CSFFW2416HS + CSFRV60J
8: CSFFW200 + CSFRV60J

2: CSFFW2416HSJ + CSFRV2216HS
6: CSFFW120 + CSFRV60J
10: 100bp



Biodynamics

AGAT

← CSFRV60J (60nt)

← CSFRV2216HS (22nt)

Amplifying with superprimers

260nt

5' ccggaggtaa aggtgtctta aagtgagaaa gaataactgc atcttaacct attgggaggt cattgtaaag aggagagtga tggggtcaga ttgtacagag gaggcacttc gtggtggtca
ggagcacaca ctccaggga gtgtccaac ctgagtctgc caaggactag caggttgcta accaccctgt gtctcagttt tcctacctgt aaaatgaaga tattaacagt aactgccttc atagatagaa
gatagataga 3'

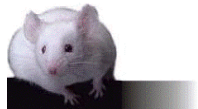
CSFFW2416HS (24nt) →

CSFFW120 (120nt) →

CSFFW200 (200nt) →

CSFFW260 (260nt) →

CSFFW300 (300nt) →



Biodynamics

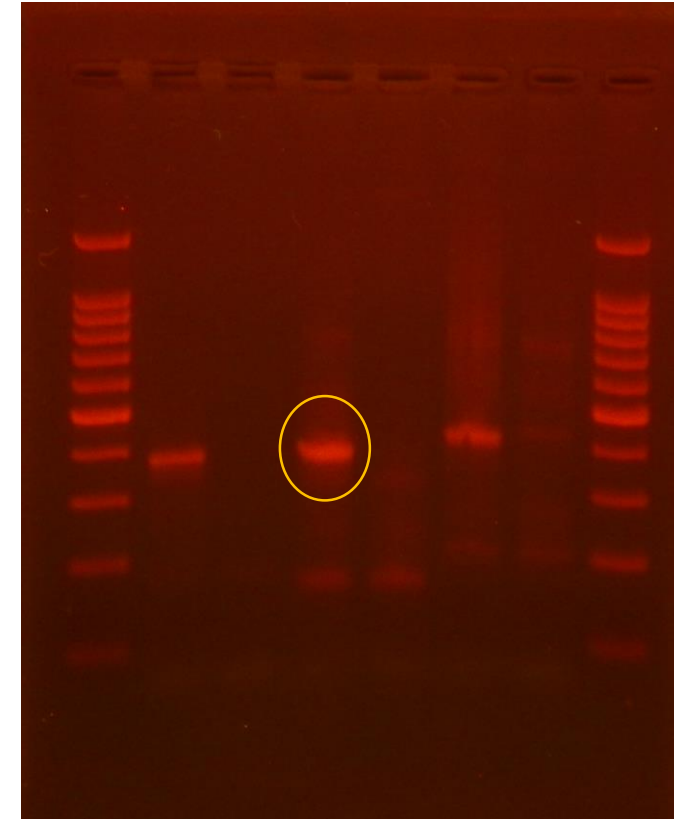
AGAT

1: 100bp
4: CSFFW260 + CSFRV60J
8: 100bp

2: CSFFW200 + CSFRV60J
6: CSFFW300 + CSFRV60J

← CSFRV60J (60nt)

← CSFRV2216HS (22nt)



Amplifying with superprimers

300nt

5' gtgaggagtg gccacagggg agcagaggag gtggcagaag ccggaggtaa aggtgtctta aagtgagaaa gaataactgc atcttaacct attgggaggt cattgtaaag aggagagtga
tggggtcaga ttgtacagag gaggcacttc gtggtgggtca ggagcacaca ctccagggca gtgttccaac ctgagtctgc caaggactag caggttgcta accaccctgt gtctcagttt tcctacctgt
aaaatgaaga tattaacagt aactgccttc atagatagaa gatagataga 3'

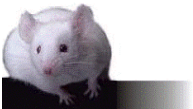
CSFFW2416HS (24nt) →

CSFFW120 (120nt) →

CSFFW200 (200nt) →

CSFFW260 (260nt) →

CSFFW300 (300nt) →



Biodynamics

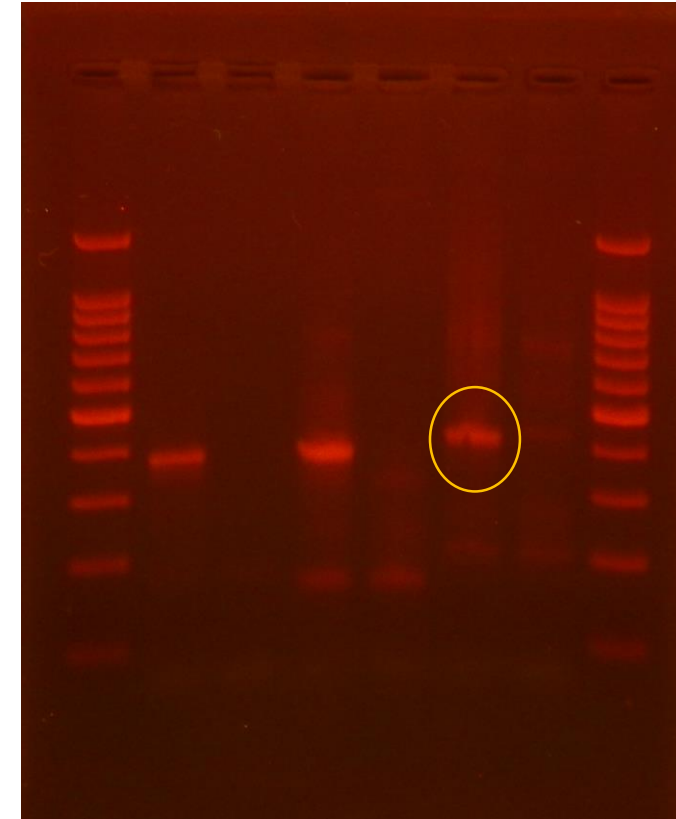
AGAT

1: 100bp
4: CSFFW260 + CSFRV60J
8: 100bp

2: CSFFW200 + CSFRV60J
6: CSFFW300 + CSFRV60J

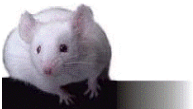
CSFRV60J (60nt)

CSFRV2216HS (22nt)



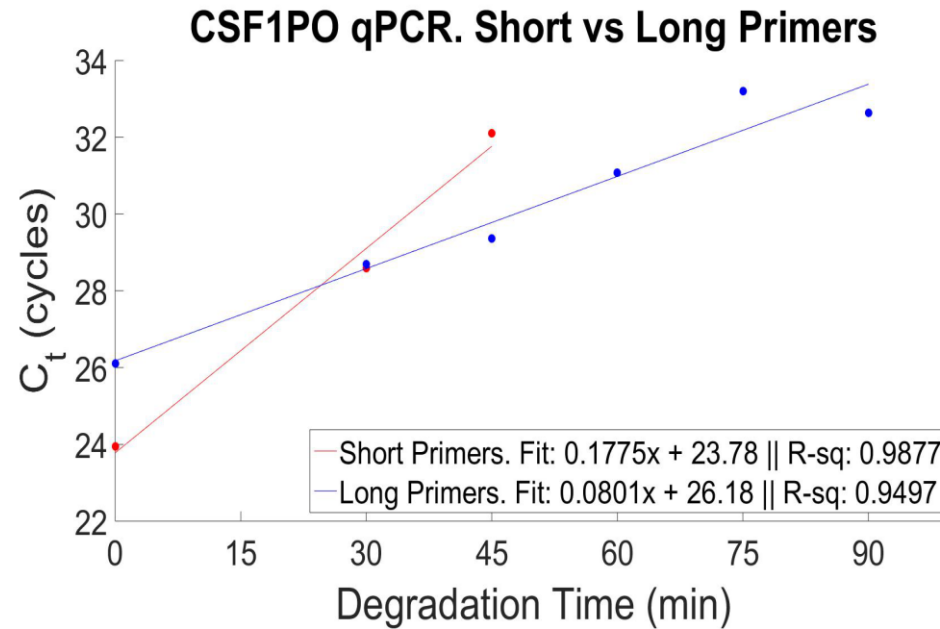
PCR conditions

- ✓ Same PCR reagents.
- ✓ Same cycling parameters.
- ✓ Same primer concentrations.
- ✓ Superprimers must be PAGE purified.



Superprimers amplify better on degraded DNA

qPCR with primer pairs CSFFW2416HS+CSFRV2216HS vs. CSFFW200+CSFRV2216HS on degraded 2800M gDNA

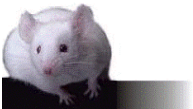


CSFFW2416HS (24nt) →

CSFFW200 (200nt) →

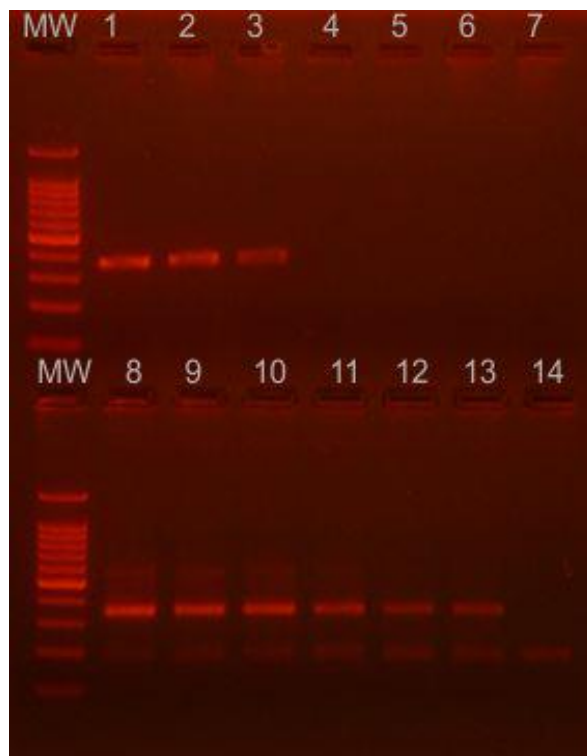
AGAT

← CSFRV2216HS (22nt)



Superprimers amplify better on degraded DNA

qPCR with primer pairs CSFFW2416HS+CSFRV2216HS vs. CSFFW200+CSFRV2216HS on degraded 2800M gDNA

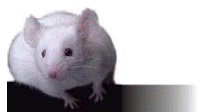


No	Heat-degradation(m in)	Primer pair	Ct
1	0	CSFFW2416HS + CSFRV2216HS	23,95
2	30	CSFFW2416HS + CSFRV2216HS	28,59
3	45	CSFFW2416HS + CSFRV2216HS	32,11
4	60	CSFFW2416HS + CSFRV2216HS	
5	75	CSFFW2416HS + CSFRV2216HS	
6	90	CSFFW2416HS + CSFRV2216HS	
7	NTC	CSFFW2416HS + CSFRV2216HS	
8	0	CSFFW200 + CSFRV2216HS	26,10
9	30	CSFFW200 + CSFRV2216HS	28,70
10	45	CSFFW200 + CSFRV2216HS	29,37
11	60	CSFFW200 + CSFRV2216HS	31,07
12	75	CSFFW200 + CSFRV2216HS	33,20
13	90	CSFFW200 + CSFRV2216HS	32,64
14	NTC	CSFFW200 + CSFRV2216HS	

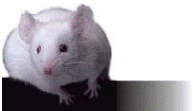
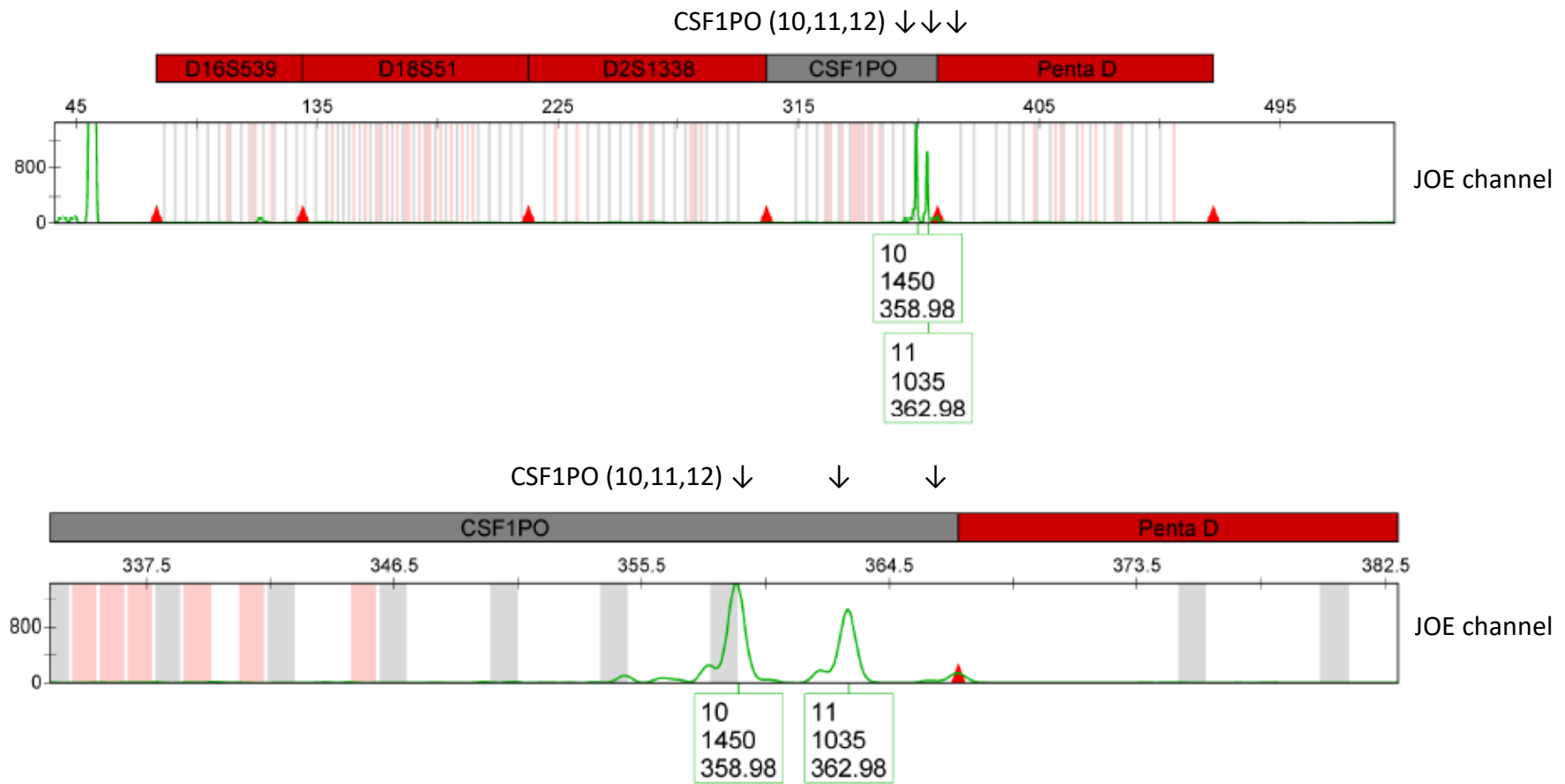
2% agarose gel electrophoresis of qPCR amplification products.

Upper panel: MW (100bp marker, Promega), samples 1-7

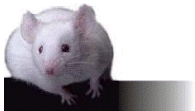
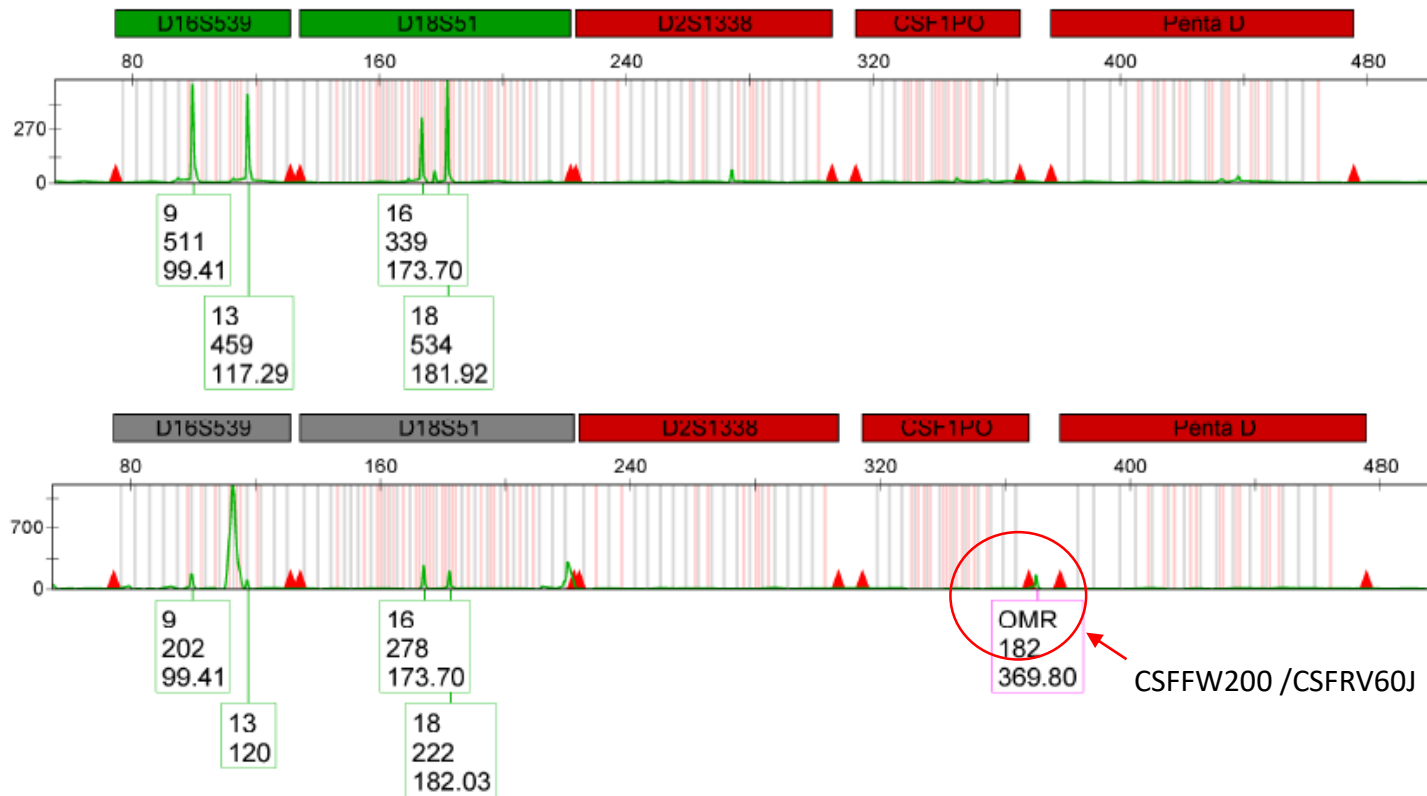
Lower panel: MW (100bp marker, Promega), samples 8-14.



Distinctive CSF1PO triplet on 9948 DNA



Spiking the superprimer pair CSFFW200 /CSFRV60J into PowerPlex™ Fusion 6C on degraded DNA (60 min)



Other STR markers

CSF1PO

CSFFW120 (120nt)



AGAT



CSFRV60J (60nt)

Penta E

PEFW120 (120nt)



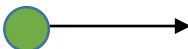
AAAGA



PERV60F (60nt)

DYS391

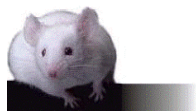
DYS391FWF (26nt)



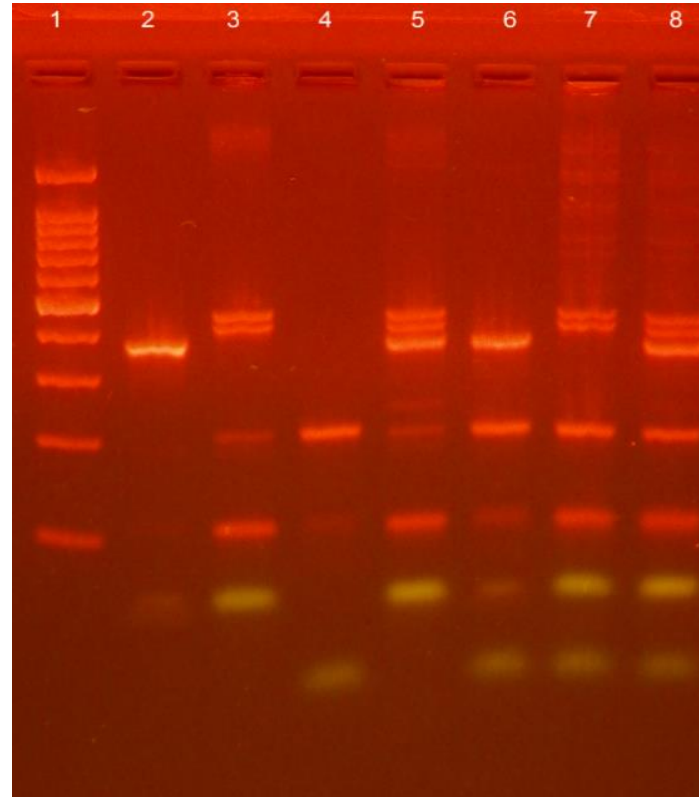
TCTA



DYSRV120 (120nt)



Multiplex PCR



1: 100bp

2: CSF1PO

3: Penta E

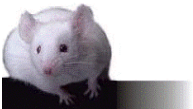
4: DYS391

5: CSF1PO + Penta E

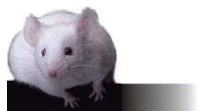
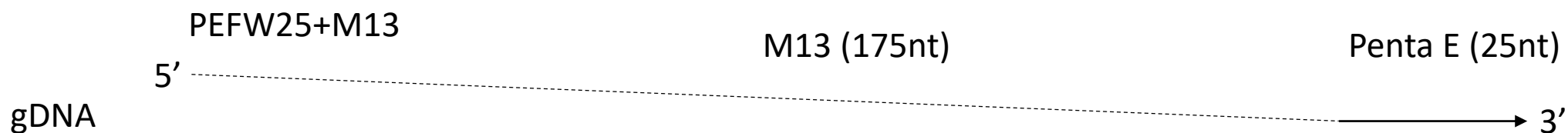
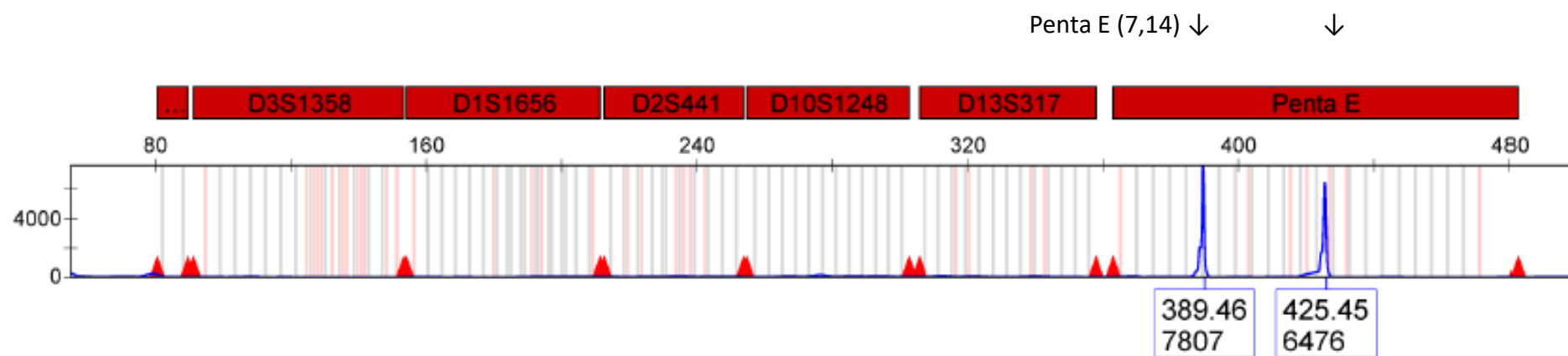
6: CSF1PO + DYS391

7: Penta E + DYS391

8: CSF1PO + Penta E + DYS391



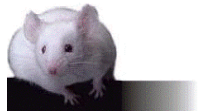
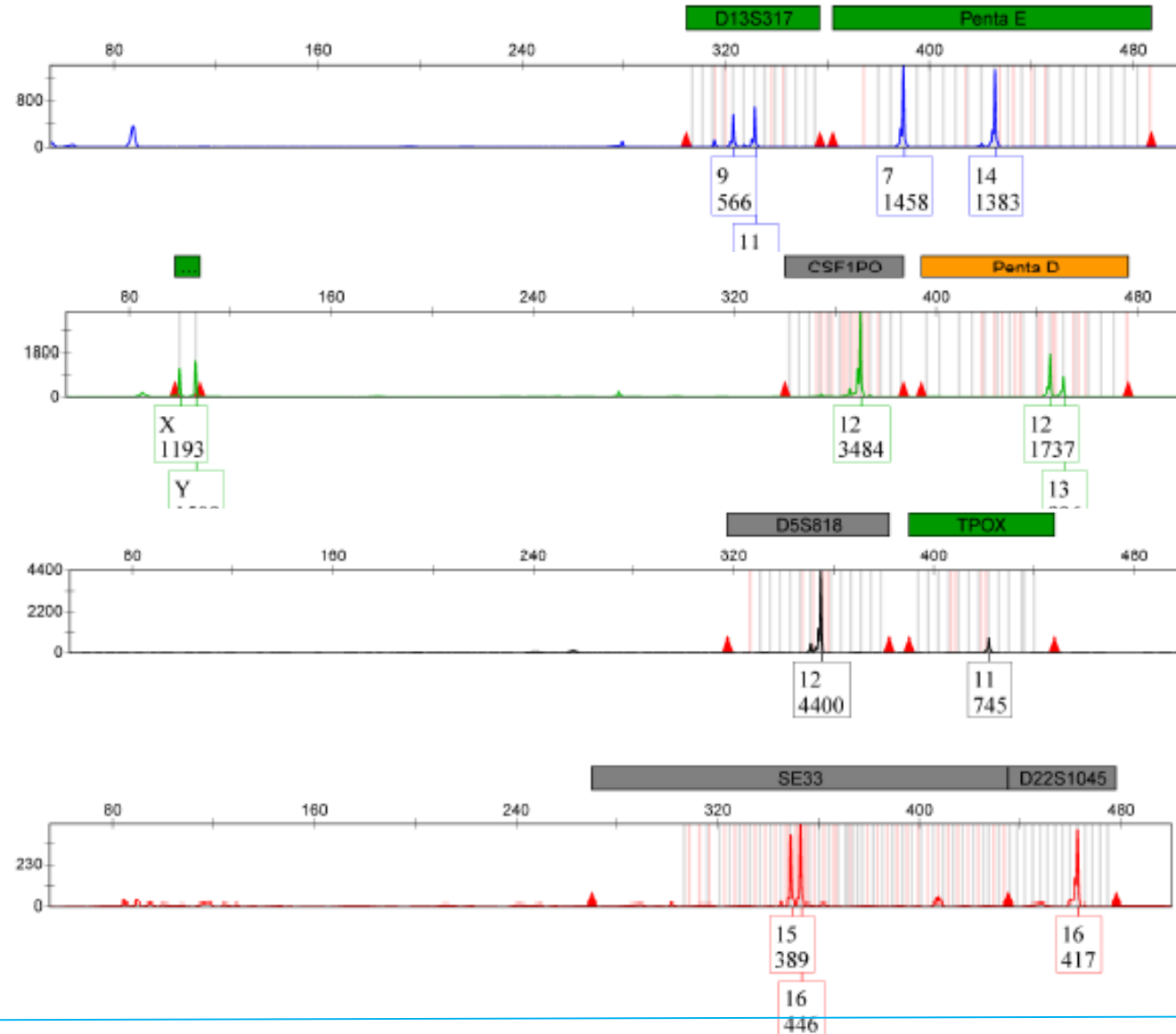
Superprimers do not need to be fully complementary



Nonaplex system

Primer name	Direction	Label	Sequence	Target locus
CSF200FW	Forward	-	cggaggtaaaggtgtcttaaagtgagaaagaataactgcatcttaacctattgggaggtcattgtaaaggagagatgatggggtcagattgtacagaggaggcacttcgtgggtggcaggagcacacactccagggcagtggtccaacctgagctgccaaggactagcaggttgctaaccaccctgtgtctcagttt	CSF1PO
CSF60RVJOE	Reverse	NHS Ester-JOE	JOE/atctcctgggtgcacacttggacagcatttcctgtgtcagaccctgttctaagtacttctct	CSF1PO
PEM1325FW	Forward	-	ccatttgcgaaatgtatctaattgggtcaactaaatctactcgttcgcagaattgggaatcaactgttacatggaatgaaacttccagacaccgtacttttagttgcatatttaaacatgttgactacagcaccagattcagcaattaagctctaagccatccgcaaaaatgaccgctgggtgtgggtgtaggcacctgt	Penta E
PE26RVFL	Reverse	56-FAM	6FAM/tgggttattaattgagaaaactccttacaattt	Penta E
D5S200FW	Forward	-	ggcttaccctcattttgaaaatacatggggagaaaaataacatagccacatttgtaattttctaattcaaaggagtatataattatgtaataattttaaattaataactgagacatgcatatgcttttaagcttctaattaaagtggtgtccagataatctgtactaataaaagtatatatttaatagcaagtatg	D5S818
D5S60RVTM	Reverse	NHS Ester-TAMRA	TAMRA/ccaatcatagccacagtttacaacatttgtatctttatctgtatccttatttatacctct	D5S818
D13M1325FW	Forward	-	acctgatttttgattatggctattctcgttttctgaactgtttaaagcatttgagggggattcaatgaatatttatgacgattccgcagtattggacgctatccagtctaaacattttactattacccctctggcaaaacttctttgcaaaagcctctcgctattttgggttttaggcagcccaaaaagacagacaga	D13S317
D13M1325RVFL	Reverse	56-FAM	6FAM/ctggtaaacgaggggtatgatagttgtgctcttaccatctaacgcctatctgtatttaca	D13S317
PDM1325FW	Forward	-	ttgctcttactatgcctcgtaattccttttggcggtatgtatctgcattagttgaatgtgggtattcctaactcaactgatgaatctttctacctgtaataatgtgttccggttagttcgttttataacgtagatttttctaaaatcgcataaggtaattcacaatgattaatacactccagcctaggtagcacagac	Penta D
PDM1325RVJOE	Reverse	NHS Ester-JOE	JOE/tcttccaacgtcctgactgggtataatgagccagttaatagggtcatgattttgtgatatc	Penta D
SE33M1329FWROX	Forward	NHS Ester-ROX	ROX/aattgatgccaccttttcagctcgcgcccaggaaggaaggaagaaaaagaaaaag	SE33
SE33200RV	Reverse	-	ccttgcgcatgctggtgcatgtgtgcagcagcagcgcggtgatagcatcatcattggtgagctggcgcggtgtcgagcgaaggcgagcggaaggacaaggttctgtgctcgtgggtgacgcggtctccggtgtgaaggaggtttatatattttctacaacatctccccaccgctatagtaacttgctc	SE33
D22S200FW	Forward	-	tccccacaggggtgactgcatctccgagtcctggcttgcacgctgacagagggtgccgagtgagcagcttaaggcatcctgccactgtgcagctgccaacctacagcccgagccctgcgggaggaagctctagtgcaggcctcttaggatctggggtccaggatgctgatttcagggccgggaccttgggcac	D22S1045
D22S60RVROX	Reverse	NHS Ester-ROX	ROX/agagtccccggcacagtgtagtgatcacgcgaatgtatgattggcaatattttataac	D22S1045
TPOX60FWTM	Forward	NHS Ester-TAMRA	TAMRA/gatcactagcaccagaaccgtcgactggcacagaacaggcacttaggaaccctcactg	TPOX
TPOX200RV	Reverse	-	actgctgtgtcgtgactccccctgtgtgggacctcgtggcggtcttacactgctgtgtcgtgactcctcctgtgtgctcacaccaagcactctcgtgtttgcacccaacgctcaaacgtgactcctcctgtactgtcctggcgctcaggggaggaactgggaaccacacaggttaattaagagattcatcaaaa	TPOX
AmelFWJOE	Forward	NHS Ester-JOE	JOE/ccctgggctctgtaaagaa	Amelogenin
AmelRV	Reverse	-	atcagagcttaaactgggaagctg	Amelogenin

Nonaplex system



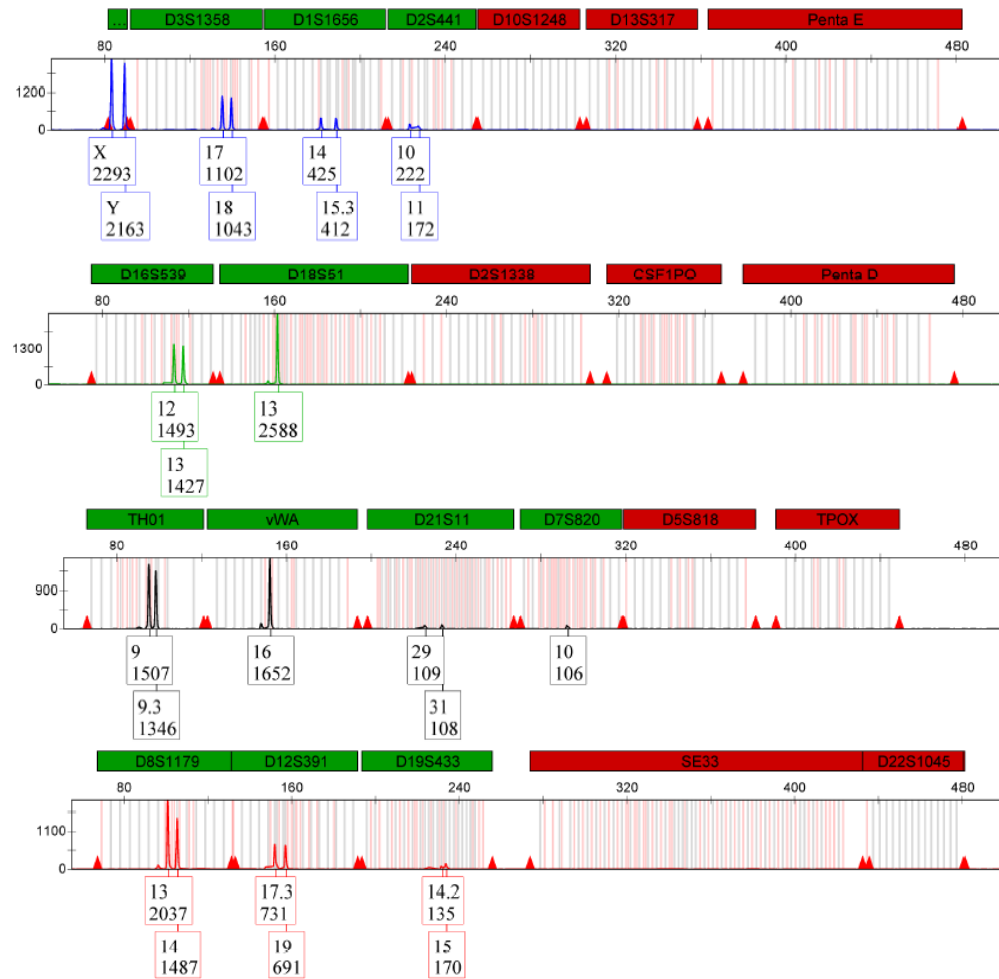
Nonaplex on cadaveric samples

Assigned genotypes for D13S317, Penta E, CSF1PO, Penta D, Amelogenin, D5S818, TPOX, SE33 and D22S1045 in cadaveric samples with PPFusion 6C vs. Nonaplex.

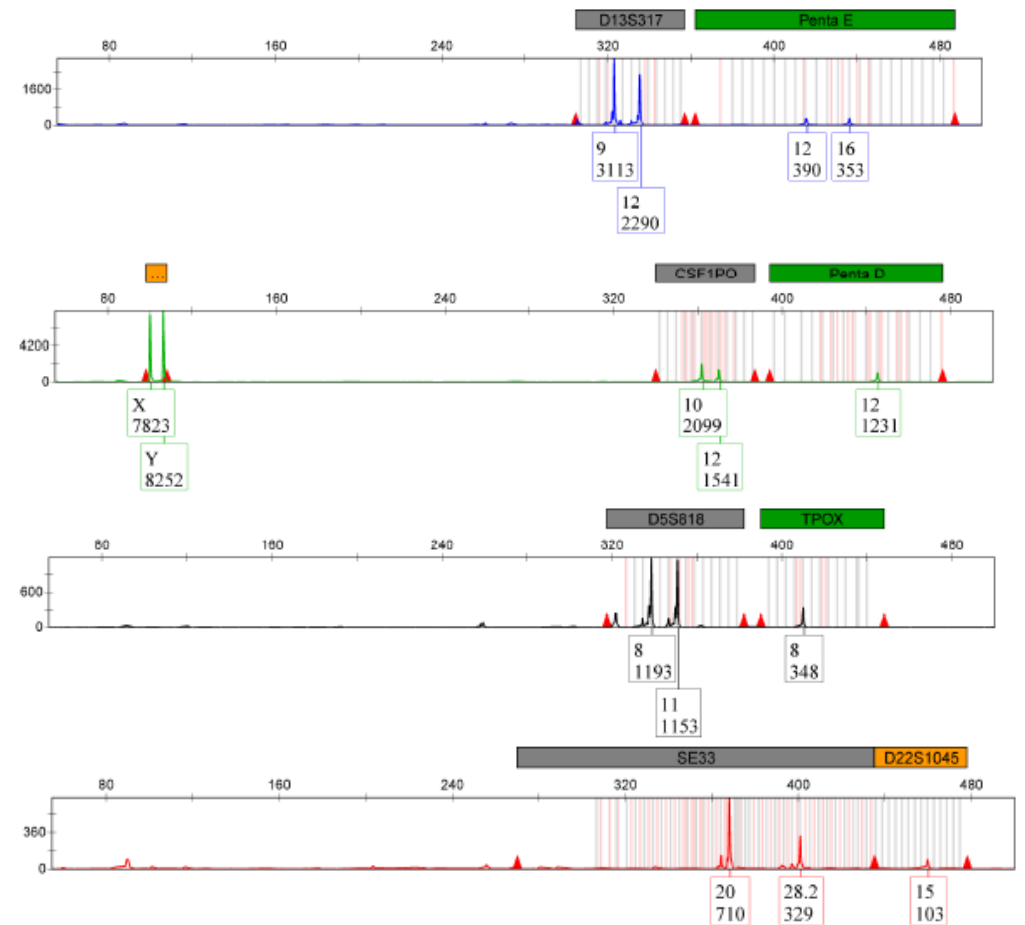
	Sample A5000		Sample A5001		Sample A5004		Sample A5163		Sample A5165		Sample A5523		Sample A5570		Sample A5616	
	Fusion 6C	Nonaplex	Fusion 6C	Nonaplex	Fusion 6C	Nonaplex	Fusion 6C	Nonaplex	Fusion 6C	Nonaplex	Fusion 6C	Nonaplex	Fusion 6C	Nonaplex	Fusion 6C	Nonaplex
D13S317	-	14	-	10	11	14	-	9,12	-	9,12	-	9,12	-	8,12	-	11
Penta E	-	11,22	-	19	-	11,22	-	12,16	-	-	-	12	-	7,10	-	7,13
CSF1PO	-	10,13	-	10,11,12	13	10,13	-	10,12	-	10,12	-	11,12	-	10,12	-	11
Penta D	-	11	-	9	-	11,13	-	12	-	-	-	10	-	-	-	9,11
Amelogenin	X,Y	X,Y	X,Y	X,Y	X,Y	X,Y	X,Y	X,Y	X,Y	X,Y	X,Y	X,Y	X,Y	X,Y	X,Y	X,Y
D5S818	-	9,11	-	11?	-	9,11	-	8,11	-	8,11	-	11	-	12,13	-	11,13
TPOX	-		-		-	8	-	8	-	-	-	8,10	-	-	-	-
SE33	-		-	-	-	19,31,2	-	20,28,2	-	20	-	17,26,2	-	22,27,2	-	16,23,2
D22S1045	-	-	-	-	-	15,16?	-	15	-	-	-	-	-	-	-	-

Nonaplex on cadaveric samples (A5163)

PPFusion 6C

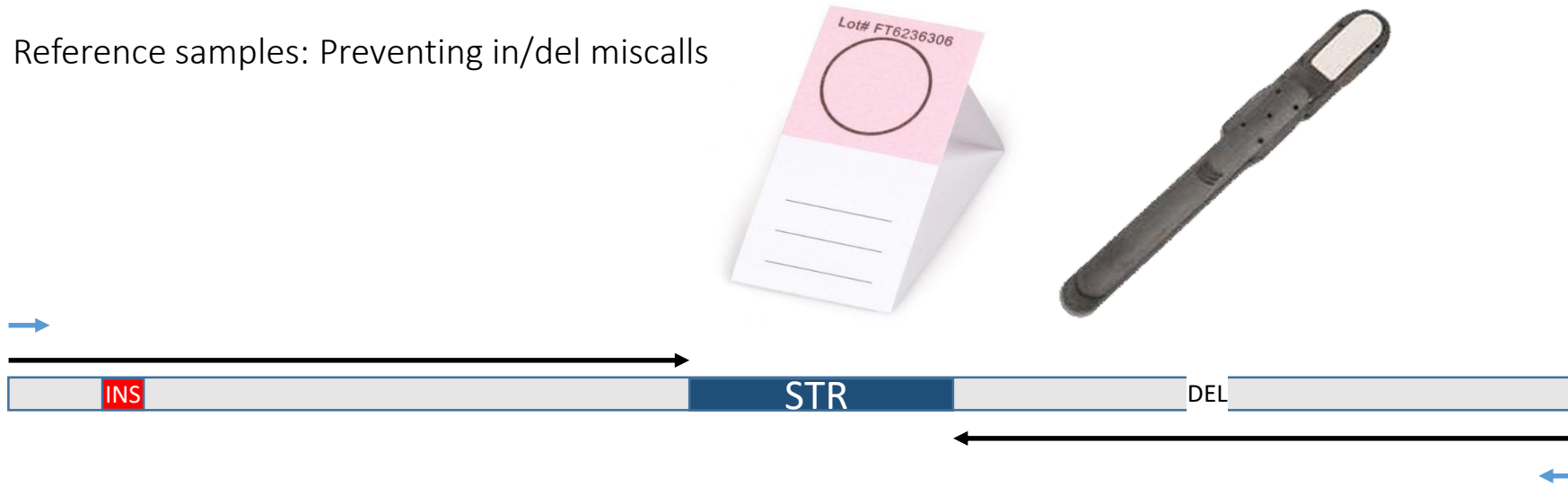


Nonaplex

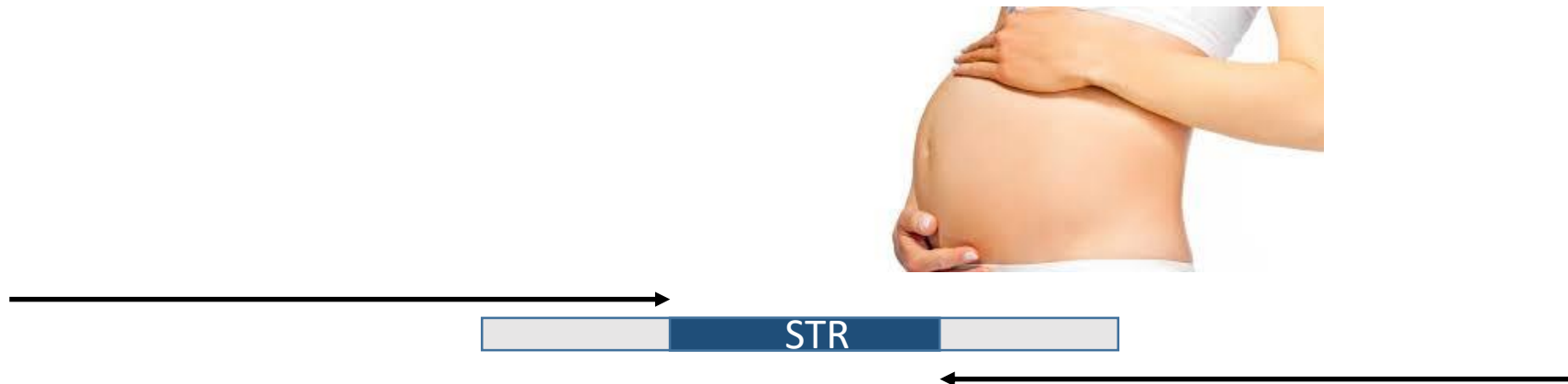


Other applications

Reference samples: Preventing in/del miscalls



Prenatal and oncogenic circulating cell-free (ccfDNA) plasma samples (170 bp)



Summary

✓ Polynucleotide primers provide more complete CE profiles for degraded DNA samples.

